Detection of Staphylococcus aureus from fish and water samples collected from Lake Qarun

Fagr Kh. Abdel-Gawad¹, Wedad E. Eweda², Gamila E. El-Taweel¹, Sawsan F. Shehata²,

Marwa I. Abdel Tawab¹.

Abstract—Staphylococcus aureus is considered to be one of the most frequently prevailing food-borne pathogen worldwide. The number of outbreaks and number of cases of staphylococcal gastroenteritis is much higher than several other microbial food borne diseases outbreaks. Staphylococcus spp. is one of the zoonotic bacteria which could be found on fish and also could reach the aquatic environment as it was isolated from different water types (fresh or brackish water) in many countries. The present study focused on detection of total staphylococci and *S.aureus* in Lake Qarun,, Egypt. Staphylococci isolates were isolated from Lake Qarun, water and Nile Tilapia (Oreochromis niloticus) fish within the period of 2010 to 2011. The determination of the four hundred typical colonies of total staphylococci and one hundred and eighty of typical colonies of *S.aureus* were carried out using surface plate technique. Moreover, molecular and biochemical confirmation of the *S.aureus* isolates were carried out by PCR analysis and biochemical reactions. Three hundred and twenty six isolates were total staphylococci +ve when tested by PCR. Also, one hundred and eighty isolates were *S.aureus* isolates +ve when confirmed by biochemical reactions.

Index Terms — Lake Qarun,, Nile Tilapia, PCR, Staphylococcus aureus.

1 INTRODUCTION

Natural lakes and reservoir surface water are a major source of freshwater for agricultural, industrial and domestic purposes worldwide. Environmental pollution, especially concerned with water sources has become of public interest. For both developed countries that have been affected and developing countries suffer from impact of pollution [1, 2, 3]. Water bodies are continually used as receptacles for untreated wastewater or poorly treated effluents increased

from industrial activities. This may provide water bodies unsuitable for both primary and/or secondary usage [4, 5].

Lake Qarun, one of the largest lakes in Egypt, is a residue of a bigger one, "Lake Moreis ", which was natively a fresh water lake. But for now, it is considered to be a closed basin for different types of wastewater drainage of EL-Fayoum governorate. Most of the discharge is carried out to the lake through two drains (EL- Batts and EL-Wadi Drain). Lake Qarun is an important source of fishing in Fayoum Provence. However, Lake Qarun, has been subjected to high level of pollution by agricultural drainage water and raw domestic sewage. Water sources contamination by wastewater is a severe health risk problem due to the abundance of pathogenic microbial agents in recreation water.

^{• (1)} Water Pollution Dep., Centre of Excellence for Advanced Sciences (CEAS), National Research Centre, Giza, Egypt

^{• (2)} Agricultural Microbiology Dep., Ain Shams University, Cairo, Egypt.

Total staphylococci or specific Staphylococcus aureus enumeration is a useful tool for water quality evaluation in different aquatic environments [6, 7].

Staphylococcus aureus determination considered as a new parameter for a long time for seawater quality evaluation. S. aureus presence in marine environments has been related to the number of bathers and may cause diseases in skin, eye or ear [8]. Also, S. aureus enterotoxins are another serious problem which causes

Water and Tilapia fish samples were collected and immediately transferred within 2-4 hours to the laboratory in NRC. Where decimal dilution method was used for detection and enumeration of staphylococci group in water samples while 3 fish samples were externally decontaminated through washing by distilled water several times then by (70%) ethanol. Fishes were dissected and 3 organs were examined for the presence of staphylococci group: liver, gills and muscle. Where 1gm of each homogenized fish organ sample was added to 9 ml of 9% saline solution then used in the experiments.

Table. 1. Lake Qarun, sampling sites description

| Site | Site description | |
|------|------------------|--|
|------|------------------|--|

1Q El-Batts drain.

- 2Q Lake Qarun, after the mixed point with El-Batts drain by 3 km.
- 3Q Lake Qarun, after the mixed point with El-Batts drain by 10 km.
- 4Q Lake Qarun, after the mixed point with El-Batts drain by 30 km.

2.2 Staphylococci group determination by surface plate technique

gastroenteritis after fish and related products consumption. Thus, this study has been conducted to detect S. aureus in water and fish samples at Lake Qarun, at different pollution sites.

2 MATERIALS AND METHODS

2.1 Samples collection

Water sample were collected monthly (from April 2010 to March 2011) from four different sites as shown in Table 1, while Tilapia fish samples were collected from 2 different sites only (1Q and 3Q).

Detection and enumeration of staphylococci group from water and Tilapia fish samples were carried out by surface plate count technique using Baird Parker agar according to APHA 2005 [9].the plates were incubated for 1-2 days at 37°C afterward five typical colonies from cultured plates were isolated and streaked on tryptic soya agar (TSA) slants and stored at 4°C.

2.3 Confirmation of total staphylococci and S. aureus by PCR:

Extractions of DNA from all samples were done as described by Kapperud et al. 1993 and Waage, A.S. et al.1999 [10, 11]. PCR assays targeting the (tuf) gene sequence has been developed for staphylococci genus as described by Martineau, F.et al.1998 [12]. The two primers used in PCR were: TStaG422 (5'-GG CC GT GT TG AA CG TG GT CA AA TC A-3') and TStag765 (5'-TI AC CA TT TC AG TA CC TT CT GG TA A-3'). and another pair of S. aureus-specific primers: Saa-442F (5'-GT CG GT AC AC GA TA TT CT TC AC G-3'), Saa442-R (5'-CT CT CG TA TG AC CA GC TT CG GT AC-3') were used to amplify 108bp which target the Sa442 gene [12].

PCR procedures were done using applied biosystems Gene Amp PCR system 9700. The PCR products were separated on 2% Agarose gel, stained with $(0.5\mu g/ml)$ ethidium bromide where a 1 kb DNA Ladder (Promega) was used as a molecular weight standard on gel as described by

Sambrook, J. et al.1989 [13]. Then gel was photographed by gel documentation system (Biometra bioanalysis).

2.4 Identification of staphylococcus aureus by biochemical reactions

Isolates were microscopically examined after Gram staining to ensure purity. Catalase, Coagulase, Sugar fermentation [14] as well as DNase production [15] tests were also performed.

3 RESULTS AND DUSCUSSION

The mixing of Qarun lake water with different types of wastewater drains including agricultural and municipal origin are heavily contaminated with various types of pathogenic microorganisms cause a severe effect on public health. A large number of epidemics due to the presence of several types of pathogens (Staphylococcus and Aeromonas bacteria) in the environment as reported by Moe, C.L.et al.2002 [16].

3.1 Density of total staphylococci and S. aureus

Results of this study were represented in Figures (1, 2 and 3) which show the density of total staphylococci and *S.aureus* in water and Tilapia fish organ samples collected from El-Batts drain and Lake Qarun, at different pollution sites using surface plate count technique were recorded during one year (April 2010 – March 2011).

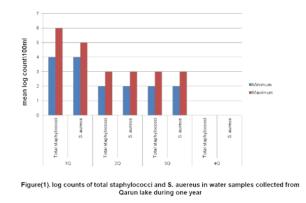
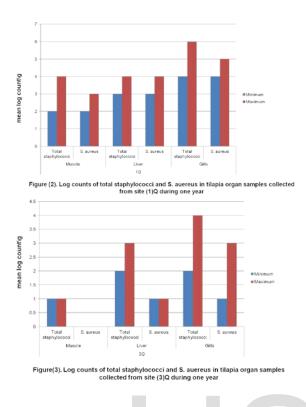


Figure (1) demonstrate the gradual increase in total staphylococci and Staphylococcus aureus density occurred after El-Batts drain is mixed with Qarun lake in sites 2Q and 3Q, with a maximum level shown in site 3Q. However, site 4Q showed no total staphylococci or staphylococcus aureus density which may be attributed to lake self purification. The above results are contradicted with study of Mansour, A.A.et al.2003 [17], who investigated some pathogenic bacteria in Lake Qarun, as there results showed that both salmonella and staphylococci bacteria were not detected in all water samples examined in Lake Qarun, at different sites. This finding may reflect the deterioration of Lake Qarun, microbial quality from 2003 till 2011.

Figure (2) and (3). Shows the density of total staphylococci and S.aureus collected from Tilapia fish organ samples. The results revealed that density of total staphylococci and Stapylococcus aureus in different fish organs are much higher in fish captured from El-Batts drain than in fish obtained from Lake Qarun,. This finding may be due to the salinity of water or because Lake Qarun, receives many effluents of drains contain industrial wastewater.



3.2 Confirmation of total staphylococci group by PCR technique

Table (2) and Figure (4) demonstrate the results of total staphylococci in water and fish samples from **El-Batts** drain and Lake Qarun,. Determination of total staphylococci was accomplished by surface plate technique then confirmed by PCR. PCR confirmation results showed that only 326 isolate was +ve staphylococci group with a percentage of 81.5%. It is important to mention that PCR failed to identify all S.aureus isolates and this may be attributed to the primer or the condition used in this test.

| Samples | Area description | | No. of | No. of +ve | |
|----------------|------------------|--------|----------|------------|----------|
| | | | isolates | | total |
| | | | tested | stap | hylococo |
| | | | | No | |
| Water | Site 1Q | | 60 | 48 | 80 |
| | Site 2Q | | 45 | 41 | 91.1 |
| | Site 3Q | | 45 | 40 | 88.8 |
| | Site 4Q | | 0 | 0 | 0 |
| Fish | Site 1Q | Muscle | 30 | 21 | 70 |
| | | Liver | 35 | 25 | 71.4 |
| | | Gills | 60 | 45 | 75 |
| Γ | Site 3Q | Muscle | 30 | 24 | 80 |
| | | Liver | 50 | 42 | 84 |
| | | Gills | 45 | 40 | 88.8 |
| Total isolates | | | 400 | 326 | 81.5 |

Table (2). The positive determination percentages of total staphylococci in water and fish organ samples.



Fig(4). Agrose gel showing results of PCR with positive control (lane 1), water (lane 2-4), fish (Lanes 5-7) isolates of Staphylococci spp and negative control (lane 8).

Marker: 100,200,300,400,500,600,700,800,900,1000,1500,3000 bp.

3.3 Confirmation of S. aureus by biochemical reactions:

Results in Table (3) showed the presence of *S.aureus* in water and Tilapia fish organ samples collected from different sites in Lake Qarun,. Also Table (4) represents the results of S.aureus isolates

IJSER © 2015 http://www.ijser.org identification by different biochemical tests. Where all *S. aureus* isolates were found to be gram positive bacteria and were able to ferment mannitol, glucose, trehalose sugar and positive to coagulase, catalase and DNase tests.

Table (3). The positive determination percentages of S.aureus in water and different fish organ samples.

| Samples | Area description | | No. of | No. of +ve | | |
|----------|------------------|--------|----------|------------|----------|--|
| | | | isolates | S.a | S.aureus | |
| | | | tested | No | % | |
| Water | 1 | Q | 20 | 20 | 100 | |
| | 2 | Q | 20 | 20 | 100 | |
| | 3 | Q | 20 | 20 | 100 | |
| | 4 | IQ | 20 | 20 | 100 | |
| Fish | | Muscle | 10 | 20 | 100 | |
| | 1Q | Liver | 20 | 20 | 100 | |
| | | Gills | 20 | 20 | 100 | |
| _ | _ | Muscle | 10 | 20 | 100 | |
| | 3Q | Liver | 20 | 20 | 100 | |
| | | Gills | 20 | 20 | 100 | |
| Total | | | 180 | 180 | 100 | |
| isolates | | | | | | |

Table (4). Identification of S.aureus isolated from water and fish samples by biochemical tests.

| B iochemical tests | | S.aureus | Others |
|-----------------------------|------------|----------|--------|
| Biochemical reaction | Gram stain | + | - |
| | Catalase | + | - |
| | Coagulase | + | - |
| Sugar fermentation | Mannitol | + | - |
| & gas production | Glucose | + | - |
| Ĵ, | Trehalose | + | - |
| DNase | | + | - |

4 Conclusion

From pervious results, it can be concluded that:

- Fish muscle contain lower or even no bacterial counts than liver and gills, while gills

contained the maximum bacterial counts especially those collected from polluted regions.

- Lake Qarun, water quality must be improved as soon as possible through prior treatment for industrial wastewater, domestic wastewater must be treated before discharge and managing agriculture wastes to reduce lake pollution.

REFERENCES

[1] A.M. Da Silva, and L.B. Sacomani, "Using chemical and physical parameters to define the quality of pardo river water (Botucatu-SP-Brazil)," Water Res., 35: 1609-1616, 2001.

[2] D. Koné, "Purification of Wastewater Lagoons Microphyte and Macrophytes in West Africa and Central: Status, Treatment Performance and Design Criteria," Ecole Polytechnique Federale de Lausanne, Lausanne, Switzerland., 7(4): 170. (in French), 2002.

[3] A. Yarar, M. Onucyildiz and N.K. Copty, "Modelling level change in lakes using neurofuzzy and artificial neural networks," J. Hydrol. 365: 329-334, 2009.

[4] R.O. Strobl, and P.D. Robillard, "Network design for water quality monitoring of surface freshwaters," A review. J. Environ. Manage., 87: 639-648, 2008.

[5] T.G. Kazi, M.B. Arain, M.K. Jamali, N. Jalbani, H.I. Afridi, R.A. Sarfraz, J.A. Baig and A.Q. Shah, "Assessment of water quality of polluted lake using multivariate statistical techniques: A case study,". Ecotoxicol. Environ. Safety, 72: 301-309, 2009.

[6] G. A. Gabutti, F. De Donno, I. Bagordon, and M. T. Montagna, "Comparitive survival of faecal and human contaminates and use of Staphylococcus aureus as an effective indicator of human pollution," Mar. Poll. Bull.,40: 697-700,2000. [7] M. M. Kamel, "Evaluation of various selective and modified media for recovery of Staphylococcus aureus from aquatic environments," Egypt. J. Appl. Sci., 20(8A): 11 – 20, 2005.

[8] L. Volterra, P. Bottoni, M. Di Carlo, and C. Dal Cero, "Balneazione: Staphylococci come indice diaollamento. L'Igiene Moderna., 101: 411-425, 1994.

[9] APHA (American Public Health Association), "Standard methods for the examination of water and wastewater," 21st ed. Washington, D.C., p.30, 2005.

[10] G. Kapperud, T. Vardund, E. Skjerve, E. Hornes and T.E. Michaelsen, "Detection of pathogenic Yersinia enterocolitica in foods and water by immunomagnetic separation, nested polymerase chain reactions and colorimetric detection of amplified DNA," Applied and Environmental Microbiol., 59: 2938-2944, 1993.

[11] A.S. Waage, T. Vardund, V. Lund and G. Kapperud, "Detection of low numbers of Salmonella in environmental water, sewage and food samples by a nested polymerase chain reaction assay," J. Applied Microbiol., 87: 418-428, 1999.

[12] F. Martineau, FJ. Picard, PH. Roy, and MG. Bergeron, "Species-specific and ubiquitous-DNAbased assays for rapid identification of Staphylococcus aureus," J. Clin. Microbiol., (36): 618–623, 1998.

[13] J. Sambrook, E.F. Fritsch and T. Maniatis, "Molecular cloning: a laboratory manual," Cold Spring Harbor Laboratory Press, New York, (2 Ed)., 3: 253, 1989.

[14] M.S. Robert, and R.K. Noel, "General characterization, chapter 20 in Manual of methods for General Bacteriology," American Society of Micribiology, Washington, DC 20006, pp: 409-433, 1981.

[15] C.D. Jeffries, D.F. Holtmann and D.G. Guse, "Rapid method for determining the activity of microorganisms on nucleic acid," J. Bact., 73: 590-591, 1957.

[16] C.L. Moe, "Waterborne transmission of infectious agents in Manual of environmental microbiology," 2 ed. ASM press, Washington, D.C., pp: 184-204, 2002.

[17] A.A. Mansour, and M.M. Sidky, "Ecotoxicological studies: heavy metals contaminating water and fish from Fayum Governorate, Egypt," Food Chem., 78: 15–22, 2003.



IJSER

372

IJSER © 2015 http://www.ijser.org